



Certificate of Mailing: Date of Deposit: September 27, 2006

I hereby certify under 37 C.F.R. § 1.8(a) that this correspondence is being deposited with the United States Postal Service as **first class mail** with sufficient postage on the date indicated above and is addressed to: Mail Stop RCE, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.

Rachel A. Kamerman

Printed name of person mailing correspondence

Signature of person mailing correspondence

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant:	Dosuk D. Lee et al.	Confirmation No.:	3866
Serial No.:	08/729,343	Art Unit:	1616
Filed:	October 16, 1996	Examiner:	Sharmila S. Gollamudi
Customer No.:	21559		
Title:	ORTHOPEDIC AND DENTAL CERAMIC IMPLANTS		

Mail Stop RCE  
Commissioner for Patents  
P.O. Box 1450  
Arlington, VA 22313-1450

DECLARATION OF DR. MICHAEL STRUNK UNDER 37 C.F.R. § 1.132

I, MICHAEL STRUNK declare:

1. I am an employee of ETEX Corporation, the owner of United States Patent Application Serial No. 10/222,670.

2. I have read and understood the Office Action, dated August 19, 2005, and Brown et al. (RE 33,221) and Simkiss (WO 94/02412), the references cited by the Examiner, and, I fully understand their technical content. This Declaration is presented to overcome the rejection of claim 25 under 35 U.S.C. § 102(b) for lack of novelty over Brown, and the rejections of claims 1, 3, 7, 9-16, and 25 under 35 U.S.C. § 103(a) for obviousness over Simkiss and Simkiss in combination with Brown, respectively.

3. I received a Ph.D. in chemistry at the University of New Mexico. I am currently employed as a chemist at ETEX Corporation. My education and professional experience are described in further detail in my curriculum vitae, a copy of which is attached as Exhibit A.

4. The present invention features a method for treating a bone defect by providing and implanting a synthetic poorly crystalline apatitic (PCA) calcium phosphate (independent claim 1) and a method for embedding a prosthetic device using a PCA calcium phosphate (independent claim 25). ETEX's PCA calcium phosphate is a carbonate hydroxyapatite, the chemical structure of which is  $\text{Ca}_a(\text{PO}_4)_b(\text{HPO}_4, \text{CO}_3)_c(\text{OH}, \text{CO}_3)_d$ , where a, b, c, and d are *non-stoichiometric* and carbonate is part of the chemical composition of the final product (see FTIR spectra, Exhibit A, enclosed). In contrast, Brown and Simkiss report a composition of  $\text{Ca}_a\text{H}_b(\text{PO}_4)_c(\text{OH})_d$ , where a, b, c, and d are *stoichiometric*. The composition of Brown and Simkiss is not a carbonate hydroxyapatite. The incorporation of  $\text{CO}_3^{2-}$  in ETEX's PCA calcium phosphate produces a change in the lattice parameters that distinguish it from the stoichiometric HA of Brown and Simkiss.

5. The Ca/P ratio of ETEX's PCA calcium phosphate is less than 1.68, which is typical for human bone. In fact, ETEX's PCA calcium phosphate is nearly identical to that of natural bone. The X-ray diffraction (XRD) pattern of ETEX's PCA calcium phosphate is similar to that of stoichiometric hydroxyapatite (HA; represented as "Clarkson HA" on Exhibit A) only to the extent that all hexagonal (H) planes of HA are represented in ETEX's PCA calcium phosphate, including the planes at  $\sim 28^\circ$  2 x theta (102, 210)(see Exhibit A). The XRD pattern provided in Fig. 1 of Simkiss does not contain all of the hexagonal planes of HA, e.g., it lacks the peaks at  $\sim 28^\circ$  2 x theta (102, 210).

6. ETEX PCA calcium phosphate is formed rapidly and directly by nucleation and growth at low temperatures (within minutes from a malleable paste) to a hardened solid; the PCA calcium phosphate is deliberately formed as a non-ordered calcium phosphate. In contrast, Simkiss reports a pre-product that precipitates from solution over a long period of time (hours), and requires the removal of inhibitor ions (i.e.,  $\text{Mg}^{2+}$ ) by leaching from the composition for hardening to occur. At hardening, the Simkiss composition converts to ordered, stoichiometric HA, not a non-stoichiometric PCA calcium phosphate, such as is recited in present claims 1, 3, 7, 9-16, and 25.

7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patents issued thereon.

Respectfully submitted,

Date: 9-26-06


  
\_\_\_\_\_  
Michael Strunk, Ph.D.



Exhibit A

Curriculum Vitae

**Michael R. Strunk, Ph.D.**

ETEX Corporation  
38 Sidney Street  
University Park at MIT  
Cambridge, MA 02139  
[mstrunk@etexcorp.com](mailto:mstrunk@etexcorp.com)

*education*

Research Fellow	Chemical Engineering University of South Carolina, 2001 – 2003 Project: <i>Non-linear Optical Spectroscopy of Heterogeneous Catalytic Interfaces</i>
M.S./Ph.D.	Chemistry University of New Mexico, 2001 / 2006 Dissertation: <i>Interface Specific Reflection-absorption Infrared Spectroscopy of Layered Solid-State Materials</i>
SRC Research Fellow	Advanced Materials & Strategic Technologies Motorola Semiconductor Products Sector (Mesa, AZ), 1998 Project: <i>In-situ crystallization Kinetics of Perovskites: Materials for Non-Volatile Memory Applications</i>
B.A.	Biochemistry University of New Mexico, 1990

*awards*

2005	Extra Mile Award, ETEX Corporation, "Accelerated Aging of Calcium Phosphate Bone Cements"
2002	1 <sup>st</sup> Place, Kristen Hedjuk, REU Mentored Project, "Biomimetic Interfaces"
1998	Smith/Dow Award, Graduate Research in Chemistry

*employment*

2004 – Current	Chemist, ETEX Corporation, Cambridge, MA
2003	Research Scientist, Visen Medical, Woburn, MA
2001–2003	Research Fellow, USC Dept of Chemical Engineering, Columbia, SC
2000	Faculty (Chemistry), Albuquerque Technical/Vocational Institute

1998–2001	Scientist, S.E.D. Medical Labs, Albuquerque, NM
1996 – 2001	Graduate Research Assistant, UNM Dept. of Chemistry
1996	Graduate Teaching Assistant, UNM Dept. of Chemistry
1990–1996	Senior Research Technologist, Chemical and Biochemical Toxicology, Lovelace Biomedical, Albuquerque, NM

#### *Selected Proceedings and Presentations*

1. M.R. Strunk, A.D. Rosenberg, M. Aioloza, S. Woods, D.C. Moore, A.S. Turner, R.D. Welch, *Comparison of  $\alpha$ -BSM<sup>®</sup> with a New, Enhanced Calcium Phosphate Cement (CaP<sub>3</sub>) in a Sheep Tibial Plateau Fracture Model*, Orthopaedic Trauma Association, 22nd Annual Meeting, Phoenix, AZ, October 2006
2. A. Hina, M. Strunk, M. Geiger, *Crystallization Kinetics of an Injectable Calcium Phosphate Based Cement*, Gordon Research Conference on Biomineralization, New London, New Hampshire, August 2006
3. M. R. Strunk, C. T. Williams. *Adsorption of aliphatic nitriles on oxide surfaces: A sum-frequency investigation*. Division of Colloid and Surface Chemistry, 226th ACS National Meeting, New York, NY, September, 2003
4. R. J. LeBlanc, I. Ortiz-Hernandez, M. R. Strunk, C. T. Williams. *In-situ vibrational spectroscopic investigation of solid-liquid catalytic interfaces*. Division of Colloid and Surface Chemistry, 225th ACS National Meeting, New Orleans, LA, March 2003
5. Strunk, M.R., Williams, C.T. *In-situ characterization of acetonitrile-Pt/Al<sub>2</sub>O<sub>3</sub> interfaces using IR-visible sum-frequency spectroscopy*, Southeastern Catalysis Society Symposium (SECS), Fall 2003
6. Strunk, M.R., Williams, C.T. *Characterization of nitrile-metal oxide interfaces using IR-visible sum-frequency spectroscopy*, American Institute of Chemical Engineers (AIChE) National Meeting, Spring 2003
7. C. T. Williams, M. R. Strunk. *Characterization of nitrile/metal oxide interfaces using IR-visible sum-frequency generation*. Division of Colloid and Surface Chemistry, 224th ACS National Meeting, Boston, MA, August 2002
8. M.R. Strunk, C.T. Williams. *Characterization of nitrile/metal oxide interfaces using IR-visible sum-frequency generation*. Gordon Research Conference on Biomineralization, New London, New Hampshire, July 2002
9. Strunk, M.R., Niemczyk, T.M. *Surface plasmon Fourier transform infrared spectroscopic investigation of a phase transition in crystalline self-assembled thiol terminated oligomers*, FACSS 2001
10. Strunk, M.R., Niemczyk, T.M. *Surface plasmon induced vibrational Stark effect spectroscopy of interfaces*, PITTCON 2000

11. Strunk, M.R., Niemczyk, T.M., Haaland, David, Clemen, Lucy *Interface sensitive multivariate modeling of reflection/absorption FTIR spectra*, FACSS 1997
12. Bechtold, W.E. Strunk, M.R. and M.J. Meyer: *S-Phenylcysteine in Albumin as a Biomarker*. Health Effects Institute Annual Meeting, Ashville, NC, April 1996
13. Henderson, R. F., W. E. Bechtold and M. R. Strunk: *Analysis of Benzene Metabolites and Glutathione in the Bone Marrow of B6C3F1 Mice*. Society of Toxicology Meeting, Baltimore, MD, March 1995.

#### *Patents*

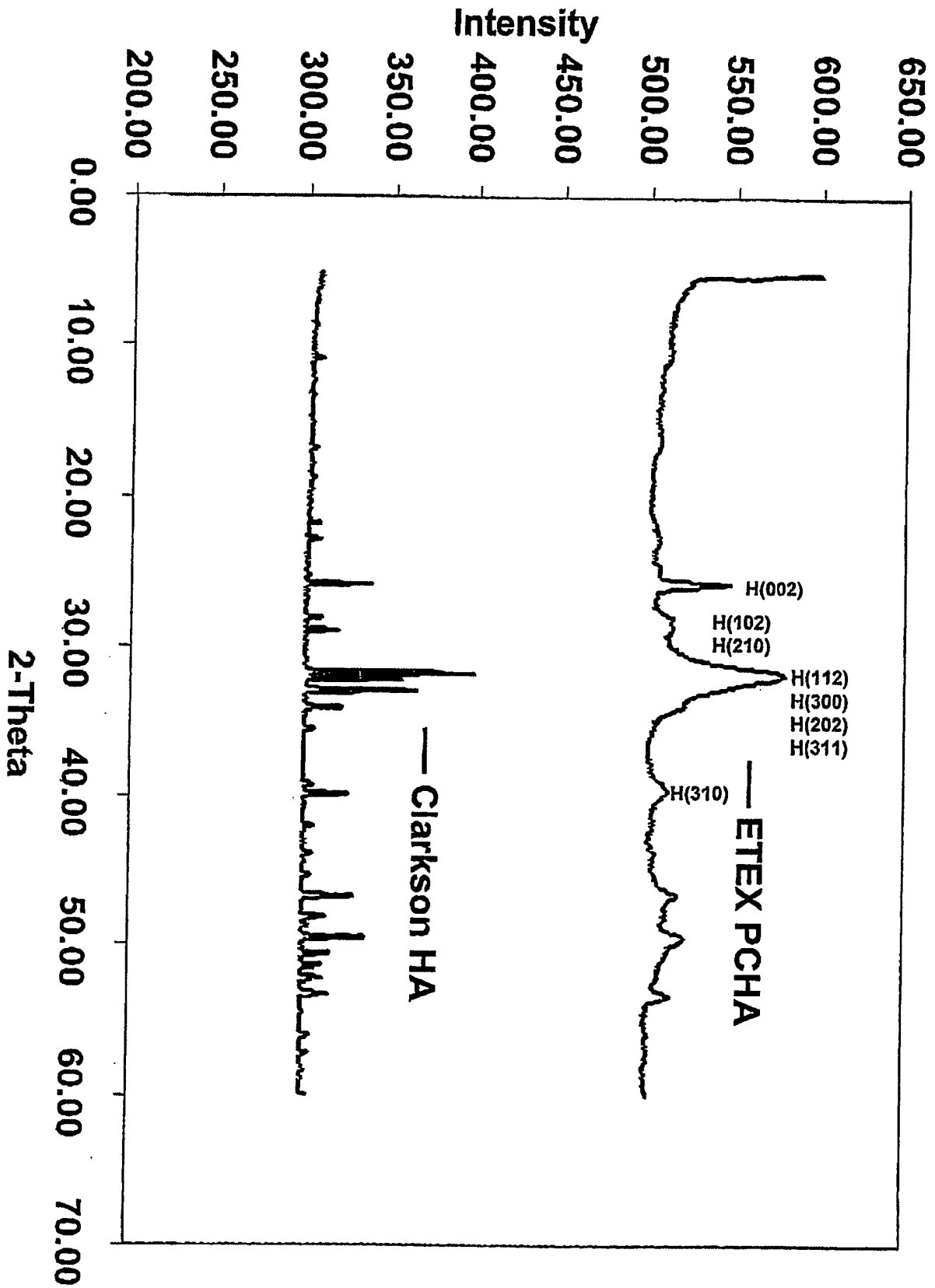
1. Rosenberg, A, Strunk, M.R., Bondre, Shrikar and Gilles de Pelichy, Laurent. Porous Calcium Phosphate Bone Cement. *Patent Filed*
2. Strunk, M.R., Niemczyk, T.M., Brueck, S.J. Interface Sensitive Vibrational Stark Effect/FTIR Spectroscopy. *provisional patent filed*.
3. Strunk, M.R. Bechtold, W.E. Automated Two-Dimensional Interface for Capillary Gas Chromatography S.N. 08/257,388 United States Patent Issued.

#### *Published Articles*

1. Ortiz-Hernandez, I.; Owens, D. J.; Strunk, M. R.; Williams, C. T. Multivariate Analysis of ATR-IR Spectroscopic Data: Applications to the Solid-Liquid Catalytic Interface *Langmuir*; 2006; 22(6); 2629-2639
2. Strunk, M.R., Williams, C.T. Aliphatic Nitrile Adsorption on Al<sub>2</sub>O<sub>3</sub> and ZrO<sub>2</sub> as Studied by Total Internal Reflection Sum-Frequency Spectroscopy. *Langmuir*, 2003
3. L.L. Clemen, M.R. Strunk, T.M. Niemczyk, D.M. Haaland Attenuated Total Reflection of Dielectric/Metal Interfaces. *11<sup>th</sup> International Conference on Fourier Transform Spectroscopy, August, 1997*
4. Pyon, K.H., Kracko, D.A., Strunk, M.R., Lewis, J.L. An Isotope Dilution Gas-Chromatography Mass Spectrometry Method for Trace Analysis of Xylene metabolites in Tissues. *Journal of Analytical Toxicology*, v.21 (#5) September 1997
5. Bechtold, W.E. Strunk, M.R., Thornton-Manning, J.R., and Henderson, R.F. Polyphenolic Metabolites in the Blood and Bone Marrow of Mice Exposed to Low-Levels of Benzene. *Abstracts of Papers of the American Chemical Society*, v. 212(pt. 1) August 1996
6. Bechtold, W.E. Strunk, M.R. S-Phenylcysteine in Albumin as a Benzene Biomarker *Environmental Health Perspectives*, v. 104(S6) Dec. 1996

7. Henderson, R. F., W. E. Bechtold, M. R. Strunk, J. R. Thornton-Manning, J. J. Waide: Analysis of Benzene Metabolites and Glutathione in the Bone Marrow of B6C3F1 Mice. *The Toxicologist* 15: 222, 1995
8. W. E. Bechtold, M. R. Strunk, J. R. Thornton-Manning, Henderson, R. F.: Benzene Metabolites Glutathione in the Blood and Bone Marrow of B6C3F1 Mice After Low Level Exposure. *ITRI Annual Report* 1994-95
9. Bechtold, W.E. Strunk, M.R., Thornton-Manning, J.R., and Henderson, R.F. Analysis of Butadiene, Butadiene Monoxide, and Butadiene Dioxide by Gas Chromatography Gas Chromatography Mass Spectroscopy. *Chemical Research in Toxicology* v8(#2) pp. 182-7 March 1995
10. Bechtold, W.E., Strunk, M.R., Chang, I.Y., Ward, J.B., Henderson, R.F. Species Differences in Urinary Metabolites: Comparisons of Ratios between Mice, Rats, and Humans. *Tox. and Applied Pharm.* 127:44-49, 1994
11. Bechtold, W.E., Strunk, M.R., Chang I.Y., Henderson, R.F, Human Metabolism of 1,3 Butadiene. *ITRI Annual Report*. 92, 1991-92
12. Bechtold, W.E., Strunk, M.R. Multidimensional HPLC Analysis of Butadiene Monoepoxide/Guanosine Adducts with Electrochemical Detection. *ITRI Annual Report*. 92, 1991-92

XRD Data





# Novel infrared spectroscopic method for the determination of crystallinity of hydroxyapatite minerals

Nancy Pleshko, Adele Boskey\* and Richard Mendelsohn

Department of Chemistry, Rutgers University, Newark College of Arts and Science, Newark, New Jersey, 07102; and

\*Department of Ultrastructural Biochemistry, Cornell University Medical College and the Hospital for Special Surgery, New York, New York, 10021 USA

**ABSTRACT** Biologically important apatite analogues have been examined by Fourier Transform Infrared Spectroscopy (FT-IR), and a method developed to quantitatively assess their crystallinity. Changes in the phosphate  $\nu_1$  and  $\nu_3$  regions, 900–1,200  $\text{cm}^{-1}$ , for a series of synthetic (containing hydroxide, fluoride, or carbonate ion) and biological apatites with crystal sizes of 100–200 Å were analyzed with curve-fitting and second derivative spectroscopy. The  $\nu_1, \nu_3$  contour was composed of three main subbands. Correlations were noted between two spectral parameters and crystal size as determined by x-ray diffraction. The percentage area of a component near 1,060  $\text{cm}^{-1}$  decreased as the length of the c-axis of the hydroxyapatite (HA) compounds increased, while the frequency of a band near 1,020  $\text{cm}^{-1}$  increased with increasing length of the apatite c-axis. These parameters are thus proposed as indices of crystallinity for biological (poorly crystalline) HA. The FT-IR spectra of highly crystalline apatitic compounds were also analyzed. For crystal sizes of 200–450 Å, the percentage area of the phosphate  $\nu_1$  band (near 960  $\text{cm}^{-1}$ ) decreased with increasing HA crystal size. IR indices of crystallinity have thus been developed for both well crystallized and poorly crystallized HA derivatives. The molecular origins of the various contributions to the  $\nu_1, \nu_3$  contour are discussed, and a preliminary application of the method to a microscopic biological sample (rat epiphyseal growth plate) is illustrated.

## INTRODUCTION

Poorly crystalline hydroxyapatite (HA) is the major mineral component of bone and other physiologically calcified tissues. Studies of HA in synthetic analogues as well as tissues have revealed many structural details (for reviews, see, for example, Bailey and Holt, 1989; Berry, 1967; Fowler, 1974a; LeGeros, 1981; Posner et al., 1979). Stoichiometric HA,  $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$  in its most common form, occurs as a hexagonally packed crystal. Apatite from biological tissues is carbonate substituted, ~5–6% by weight (LeGeros, 1981). The  $\text{CO}_3^{2-}$  ion can substitute for either the phosphate or the hydroxyl groups of HA, leading to a nonstoichiometric apatite mineral (Bonel, 1972a, b; Elliot, 1961; Emerson and Fischer, 1962). Other possible substitutions include replacement of the hydroxyl group by fluoride ion. This may be an important event during the treatment of the bone disease osteoporosis with NaF (Gron et al., 1966).

The structure of HA analogues have been extensively studied by various chemical and physical methods (Blumenthal et al., 1975; Eanes et al., 1973; Fowler et al., 1966; Posner et al., 1979; Termine and Lundy, 1973). In particular, the crystallinity, i.e., crystal size and perfection, of apatites are of great biomedical interest, because smaller, more imperfect crystals, being subject to dissolution, may effect the extent of bone loss in osteoporosis and other metabolic bone diseases (Thompson et al., 1983). Traditionally, x-ray diffraction is the technique of choice for determination of apatite crystal size. The method, while direct, cannot be applied to microscopic

samples with rapid spatial variation in mineral structure such as biological tissue specimens.

In contrast, structural information relating to the apatite mineral in macroscopic, as well as microscopic samples, can be readily obtained from IR spectroscopy. Termine and Posner (1966) developed an IR method to determine the percentage of crystallinity in apatite minerals, based on changes in the phosphate  $\nu_4$  mode. Others have carried out analyses of calcium phosphates to determine the molecular species contributing to the characteristic phosphate bands (Baddiel and Berry, 1966; Bhatnagar, 1988; Cant et al., 1971; Fowler et al., 1966; Fowler, 1974a, b; Stutman et al., 1965). Fourier deconvolution techniques have recently been used to analyze spectra of both synthetic and biological calcium phosphates (Rey et al., 1991; Rey et al., 1990).

This study describes a novel IR method to assess the crystallinity of HA minerals, based on changes in the phosphate  $\nu_1$  and  $\nu_3$  absorbances in the 900–1,200- $\text{cm}^{-1}$  spectral region. The technique is suited to both macro- and microscopic samples, and serves to complement x-ray diffraction analysis of crystal size and perfection. The method, developed from spectra of biologically-relevant hydroxyl, carbonate, and fluoride-substituted apatites of known crystal sizes, was then employed to determine the crystal size of apatite in rat femur epiphyseal growth plates, a problem not readily addressed by x-ray diffraction techniques.

## EXPERIMENTAL

### Materials

Poorly crystalline HA was prepared from amorphous calcium phosphate (ACP) in 0.15 M Tris buffer at room temperature, pH 8.5 (Boskey and Posner, 1973). Carbonate-substituted HA was prepared from ACP in a solution containing 0.15 M Tris and 0.026 M  $\text{Ca}_3\text{CO}_3$  at pH 8.5 or 7.4, (Blumenthal et al., 1975). Fluoride-substituted HA was prepared by the addition of NaF to ACP in solution, pH 8.5 (Hasvold and Dahm, 1977). Ca/F mole ratios of 10 to 1, and 5 to 1, respectively, were used. For each reaction mixture, aliquots of the slurry were removed at sequential time points (ranging from time "0", start of the reaction, to 3 wk), filtered, washed with pH 10 water, and dried with acetone. The precipitate was dried under vacuum overnight, and stored desiccated until use. KBr pellets of the precipitates were prepared using 0.5 mg sample/100 mg KBr, and their FT-IR spectra recorded. The carbonate content of the compounds was determined by the Conway microdiffusion method (Conway, 1957), and the fluoride content determined by measurement with a fluoride specific ion electrode.

Well crystallized apatites examined included geological fluorapatite and chlorapatite, hydroxyapatite prepared by refluxing, and carbonate substituted apatite prepared at 37°C (kindly supplied by Dr. R. LeGeros, New York University School of Dental Medicine, New York).

Bone samples were ground in a liquid nitrogen cooled colloid mill (Spex Industries, Metuchen, NJ), and sieved through a metal 325  $\mu$  mesh. KBr pellets for IR examination of ground bone were prepared using 0.5 mg sample/100 mg KBr.

Thin (5  $\mu$ ) rat femur sections were cut on a Tissue-Tek No. 2 Cryostat (Electrobiology, Inc., Fairfield, NJ) equipped with an American Optical steel microtome knife.

### Methods

**FT-IR spectroscopy.** FT-IR spectra were recorded with a SIRIUS 100 spectrophotometer (Mattson Instruments, Inc., Madison, WI), equipped with a Bach-Shearer FT-IR microscope and a Mercury-Cadmium-Telluride detector. Routinely, 400 interferograms were collected, coadded, apodized with a triangular function, and fast Fourier transformed to yield spectra with 4  $\text{cm}^{-1}$  resolution with data encoded every 2  $\text{cm}^{-1}$ .

**X-ray diffraction.** The structure of the apatites were confirmed by wide angle x-ray diffraction, using Cu K- $\alpha$  radiation on a Siemens automated powder diffractometer (Siemens-Allis Inc., Cherry Hill, NJ). Crystallinity was assessed from the line broadening of the 002 (*c*-axis) reflection.

### Data analysis

Crystal sizes of the apatites were calculated from x-ray diffraction line width data. The width at half height of the 002 reflection,  $\beta_{002}$ , is inversely proportional to the crystallite length along the *c* unit cell direction (Cullity, 1956), which is termed the "*c*-axis length" in this study, as follows.

$$D = \frac{0.9\lambda}{\beta_{002} \cos\theta}$$

where *D* is the crystal size ( $\text{\AA}$ );  $\lambda$  is the wavelength of the incident x-rays, 1.54  $\text{\AA}$ ;  $\beta_{002}$  is the width at half height of the 002 reflection; and  $\cos\theta$  is the cosine of the x-ray incident angle (25.85°).

The 900–1200- $\text{cm}^{-1}$  IR spectral region contains the phosphate symmetric and antisymmetric stretching vibration,  $\nu_1$  and  $\nu_3$ , respec-

tively. Analysis of the contour was carried out with software developed at the National Research Council of Canada, Ottawa, and generously supplied by D. Moffatt. The curve-fitting algorithm creates Lorentzian-Gaussian bands that are added to produce a computed spectrum, which is compared with the experimental spectrum. The process is iterated until a satisfactory fit between the computed and experimental band is obtained by a least squares regression analysis. The calculated area of each subband is reported as a percentage of the computed contour. Initial peak positions were obtained from second-derivative spectra of the raw data (Fig. 1A). The relative proportions of Lorentzian-Gaussian bandshapes are varied for the set of subbands during the iteration.

Because the analysis depends on the positions and intensities of the components of the contour, substantial attention was paid to obtaining the best possible fit with the minimum number of component bands. It was evident from the second derivative spectra (see Fig. 1A for an example) that at least four bands were required. Initial iterations produced obvious difference patterns which guided the selection of the remaining two features. The effect of the number of bands on the

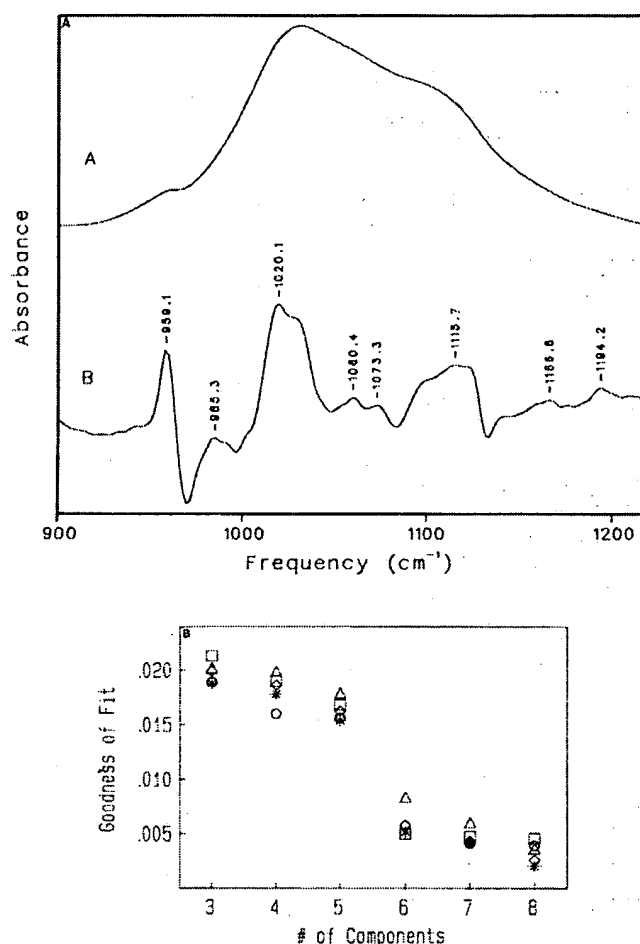


FIGURE 1 (A) Typical FT-IR spectrum of poorly crystalline hydroxyapatite, phosphate  $\nu_1, \nu_3$  region. Second derivative spectrum (B) shows at least four underlying components. (C) Goodness of fit versus number of components used in the curve fit analysis of poorly crystalline carbonate-substituted apatites. Circle, 90 min in solution; square, 3 h; asterisk, 24 h; diamond, 72 h; triangle, 3 wk.

quality of the fit for five separate samples is shown in Fig. 1 *B*. In each case, the goodness-of-fit (GOF) parameter from the least squares algorithm (the normalized root mean square deviation), was determined as a function of the number of bands used for the fit. In all cases, the improvement in GOF diminishes drastically with addition of a seventh component (Fig. 1 *B*); it was thus evident (and consistent results for more than 100 data sets were obtained using this criterion) that six bands were both necessary and sufficient to accurately represent the contour of poorly crystalline apatites.

## RESULTS

Spectral changes accompanying HA crystal growth were evaluated during the solution-mediated transformation of amorphous calcium phosphate (ACP),  $\text{Ca}_3(\text{PO}_4)_2 \cdot \text{XH}_2\text{O}$ , to HA,  $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$ . ACP undergoes an autocatalytic reaction in aqueous solution to form a poorly crystalline HA (Boskey and Posner, 1973), resulting in crystals ranging in size from 100 to 200 Å. As the HA crystals mature, the IR spectra of the calcium phosphate precipitates taken at sequential time points are substantially altered (Fig. 2). Qualitatively, the shape of the  $\nu_1, \nu_3$  absorbance progressively changes from the broad, generally featureless (except for the shoulder at 960  $\text{cm}^{-1}$ ) contour seen in ACP, to a composite of overlapping narrower bands from which a high frequency shoulder eventually emerges. The broad  $\nu_4$  absorbance band in the 500–700- $\text{cm}^{-1}$  region also becomes resolved into two distinct peaks, characteristic of

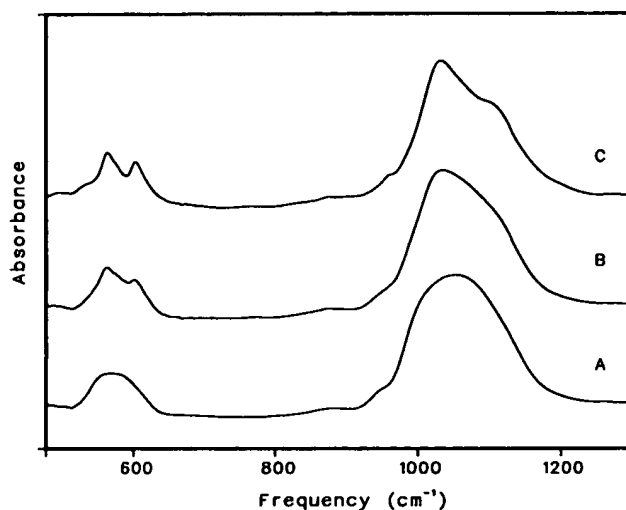


FIGURE 2 FT-IR spectra showing the solution-mediated, autocatalytic conversion of amorphous calcium phosphate (ACP) to poorly crystalline HA. *A*: ACP; *B*: HA, in solution 90 min; *C*: HA, 24 h in solution. With time, the phosphate  $\nu_1, \nu_3$  region (900–1,200  $\text{cm}^{-1}$ ) changes from a broad band, as in ACP, to a narrower band with a high frequency shoulder. The  $\nu_4$  region (500–700  $\text{cm}^{-1}$ ) becomes resolved into two distinct peaks.

hydroxyapatite. As noted above, we have correlated changes in the 900–1,200- $\text{cm}^{-1}$  region with the size (c unit cell dimension) of the HA crystals.

Because biological apatites contain ~5–6% carbonate by weight, it was deemed useful to include carbonate-substituted apatites in this study. In addition, the widespread use of fluoride as a therapeutic agent in the treatment of osteoporosis (Baud et al., 1988; Gron et al., 1966; Kragstrup et al., 1989), led us to examine fluoride-substituted materials.

The spectral features between 900 and 1,200  $\text{cm}^{-1}$  arise primarily from the symmetric ( $\nu_1$ ) and antisymmetric ( $\nu_3$ ) P-O stretching modes of the ACP and HA phosphate groups. Under strict tetrahedral symmetry for orthophosphate,  $\nu_1$  is IR inactive. However, when the symmetry in the crystal is lowered from that of the free ion (e.g., in HA), the symmetric stretch appears as a weak feature in the IR spectra between 950–970  $\text{cm}^{-1}$ . The  $\nu_3$  mode is triply degenerate in the free ion, and may resolve into three separate modes when the degeneracy is lifted. We therefore expect that analysis of the 900–1,200- $\text{cm}^{-1}$  region would lead to multiple bands arising from both the phosphate  $\nu_1$  and  $\nu_3$  modes.

Bands in the 500–700- $\text{cm}^{-1}$  region arise primarily from the antisymmetric P-O bending modes of the phosphate groups. This triply degenerate mode ( $F_2$  in  $T_d$  symmetry), is resolved into at least two well defined peaks in HA (Baddiel and Berry, 1966). Analysis of this spectral region produced no conclusive spectra-structure correlations.

The weak absorbance band at ~870  $\text{cm}^{-1}$  arises primarily from  $\nu_2$  ( $A_1'$ ) of carbonate (Termine and Lundy, 1973), which may be incorporated into the apatites from atmospheric  $\text{CO}_2$ . Without addition of  $\text{CaCO}_3$  to the ACP reaction mixture, the carbonate content of the HA was ~2% by weight after 3 wk in solution.

Fig. 3 displays spectra representative of poorly crystalline HA, with the experimental and calculated contours overlaid along with the individual subbands. Six components were necessary and sufficient for a suitable fit in the 900–1,200- $\text{cm}^{-1}$  spectral region, with the three main features near 1,020, 1,050, and 1,110  $\text{cm}^{-1}$  being termed the A, B, and C marker bands, respectively. Similar results were obtained for carbonate and fluoride substituted apatites, respectively (Fig. 4). The position, intensity, and linewidth of each component were examined. Correlations were noted between two of the parameters (the fractional intensity of the B band, and the position of the A band) with crystal size of the apatites.

The principal relationship was a decrease in the percentage area of component B, from ~45 to 5%, as the crystal size of the apatites increased from ~135 to 190 Å (Fig. 5 *A*). From these data it is clear that a

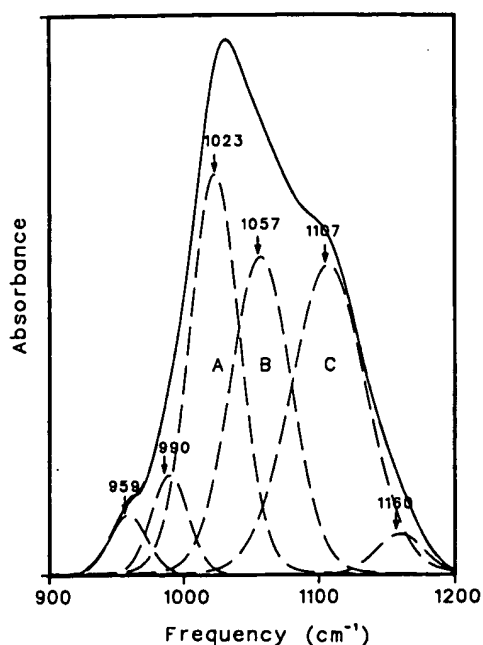


FIGURE 3 Typical FT-IR spectrum of poorly crystalline HA, phosphate  $\nu_1, \nu_3$  region (—), with an overlay of the calculated contour, and the individual components (----) as determined by a curve fitting analysis. Six components were needed for a satisfactory fit, of which components A, B, and C were the main features of the contour.

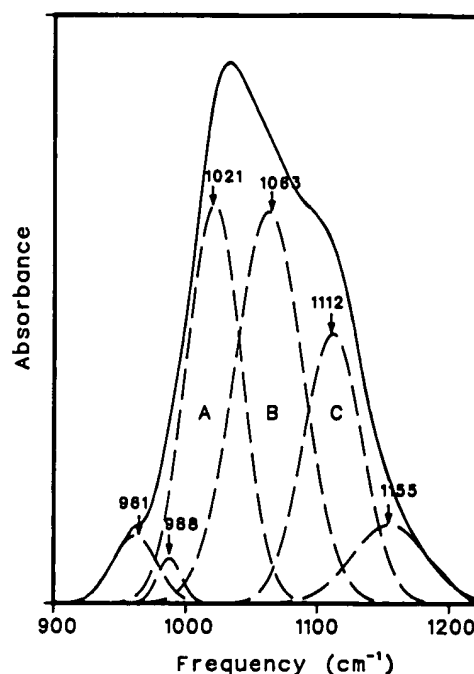


FIGURE 4 Typical FT-IR spectrum of a carbonate-substituted HA (4.5% carbonate by weight), phosphate  $\nu_1, \nu_3$  region (—), with an overlay of the calculated contour, and the individual components (----) as determined by a curve fitting analysis.

different spectral parameter is required for crystal sizes  $> 200 \text{ \AA}$ , because as the area of the B band is diminished in intensity, it becomes too difficult to measure accurately. However, for apatites with sizes in this range, knowledge of the percentage area of component B gives a useful indication of crystal size. Thus, an index of crystallinity of poorly crystalline HA compounds, regardless of atomic (chemical) substitution, has been developed.

To verify that the index of crystallinity is applicable to biological materials, FT-IR spectra were collected from KBr pellets of four different ground bone samples, including rat, bovine, and rabbit bone (whose principal mineral component in each instance is a poorly crystalline HA [LeGeros, 1981]). The  $900\text{--}1,200\text{-cm}^{-1}$  region for each spectrum was curve fit, and the percentage area component B versus the crystal size datum for each sample is included in Fig. 5A. The data agree extremely well with those from the model compounds, demonstrating that the correlation extends to biological samples. Nonlinear regression analysis of the data in Fig. 5A led to the following relationship between fractional area of the B band and  $c$ -axis crystal size:

$$Y = aX^b + c \quad (r = 0.84)$$

$$Y = \text{Crystal size (\AA)}$$

$X = \text{Percentage area component B}$

$$a = -30.70$$

$$b = 0.2840$$

$$c = 227.8. \quad (1)$$

The quantitative correlation proposed above is valid for interpolation purposes for  $c$  unit cell lengths between 195 and 135  $\text{\AA}$ . A second semiquantitative relationship is apparent between the frequency of the A component and the crystal size of the apatites. The frequency is not greater than  $1,027\text{-cm}^{-1}$  for apatite crystal sizes less than  $\sim 160 \text{ \AA}$ , and increases to  $1,028\text{--}1,032\text{-cm}^{-1}$  for crystal sizes between  $\sim 160\text{--}190 \text{ \AA}$ . This parameter is thus appropriate as a secondary indicator of crystal size for poorly crystalline specimens. The remaining four components of the contour could not be related to HA crystal size.

Well crystallized apatites ( $200\text{--}450 \text{ \AA}$  in size) revealed additional spectral features compared with the more poorly crystalline materials. Fig. 6 shows a spectrum typical of a well crystallized apatite in the  $900\text{--}1,200\text{-cm}^{-1}$  region. Qualitatively, the central contour is considerably narrower than that of the poorly crystalline HA, with the high frequency shoulder more clearly defined. The band requires at least eight components for an

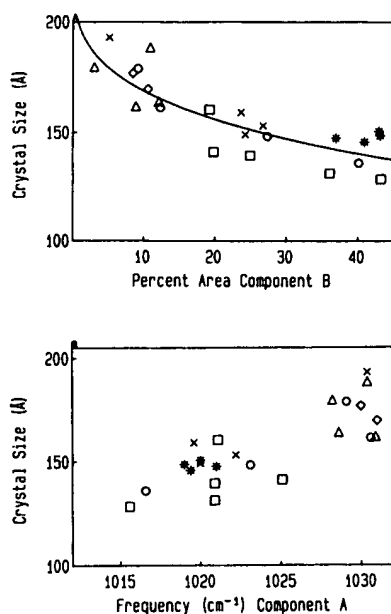


FIGURE 5 (A) Relationship of crystal size ( $\text{\AA}$ ) (c-axis dimension) to percentage area component B, and (B) to the position of component A (see Fig. 3) as determined by curve fitting analysis, for a series of poorly crystalline apatite analogues in the 100–200  $\text{\AA}$  range. HA, prepared at pH 8.5 ( $\circ$ ); carbonate-substituted HA, prepared at pH 8.5 ( $\square$ ); carbonate-substituted HA, prepared at pH 7.4 ( $*$ ); fluoride-substituted HA, prepared at pH 8.5 ( $\diamond$ ); fluoride-substituted HA, prepared at pH 7.4 ( $\triangle$ ); biological samples, as discussed in the Experimental section ( $x$ ).

adequate fit to the spectrum, compared with only six in the less crystalline preparations. Due to the additional features, it was not realistic to attempt to identify particular components characteristic of mineral size in this series of materials. However, a correlation (Fig. 7) was found between the percentage area of the  $\nu_1$  band (near 960  $\text{cm}^{-1}$ ) and crystal size. The  $\nu_1$  component decreases from  $\sim 1.9$  to 0.5% as crystal size increases from  $\sim 200$  to 450  $\text{\AA}$ . This constitutes a useful index of crystallinity for well crystallized apatites.

We note that it is easy to determine which type of apatite, poorly or well crystallized, one is analyzing, and therefore which of the available spectral indices are appropriate for crystal size determination. The contour in the 900–1,200- $\text{cm}^{-1}$  spectral region is much narrower for the well crystallized apatites (crystal size  $> 200$   $\text{\AA}$ ), and the  $\nu_1$  component is clearly resolved. Biological apatites generally have crystal sizes comparable to the poorly crystalline preparations examined in this study, and therefore the “A” and “B” components derived from the data analysis yield the appropriate indices of crystallinity.

The utility of this method for HA crystal size determination is demonstrated through FT-IR microscopy stud-

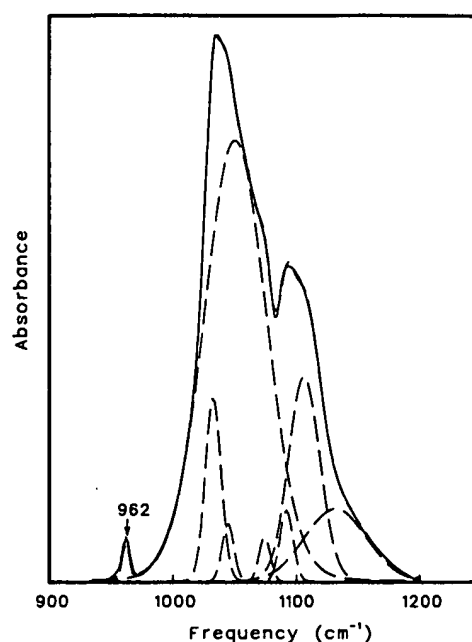


FIGURE 6 Typical FT-IR spectrum of a well crystallized apatite, HA prepared by refluxing for 24 h (—), with an overlay of the calculated contour, and the individual components (---), as determined by a curve-fitting analysis. Eight components were needed for a satisfactory fit.

ies of thin (5  $\mu$ ), sagittal rat femur sections. FT-IR spectra were collected from discrete areas, 30  $\mu$  in diameter, of the femur epiphyseal growth plate, where the newest mineral is formed (Warshawsky, 1982). Examination of the 900–1,200- $\text{cm}^{-1}$  spectral regions reveal obvious qualitative differences in the contour of this band, corresponding to distinct regions of the growth plate. Application of the method described above for crystal size determination led to quantitation of the apatite mineral dimension at various regions of

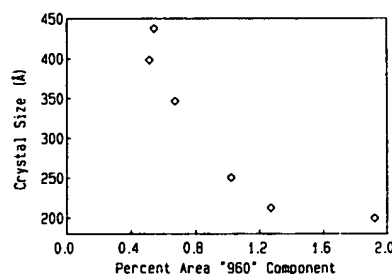


FIGURE 7 Relationship of crystal size ( $\text{\AA}$ ) (c-axis dimension) to percentage area  $\nu_1$  component as determined by curve-fitting analysis, for a series of well crystallized apatite analogues, including fluorapatite, chlorapatite, apatite prepared by refluxing, and carbonate-substituted apatites prepared at 37°C.

growth plate (Fig. 8 A, B). Although detailed interpretation of the biomedical and developmental significance of these results will be deferred for a more appropriate forum, it is clear that we have obtained information about the crystallinity of the mineral at 30  $\mu$  spatial resolution, an experiment not feasible for conventional x-ray diffraction techniques.

## DISCUSSION

A spectroscopic method has been developed to assess the crystallinity of hydroxyapatite minerals. The observed changes in the 900–1,200- $\text{cm}^{-1}$  spectral region, which reflect the maturation of HA crystals, have been quantitated by a least squares curve fitting analysis. Six components were needed to obtain a satisfactory fit for the poorly crystalline apatites studied (125–190  $\text{\AA}$ ), whereas seven or eight components were needed to fit the well crystallized apatite (200–450  $\text{\AA}$  in size) spectra. A discussion of the possible molecular origin of the components follows.

The phosphate ion,  $\text{PO}_4^{3-}$ , is the principal molecular species giving rise to the HA absorbance in the 900–1,200- $\text{cm}^{-1}$  region. Fowler et al. (1966) have characterized the infrared frequencies of HA. The  $\nu_1$  absorbance occurs at 962  $\text{cm}^{-1}$ , while two components of  $\nu_3$ , at 1,040 and 1,092  $\text{cm}^{-1}$ , were identified.

The relationship demonstrated here between two components of the  $\nu_1, \nu_3$  contour, and the crystal size of the poorly crystalline apatite minerals may be used to aid in the assignment of the phosphate ion bands. The percentage area of component B, centered near 1,060  $\text{cm}^{-1}$ , was shown to decrease as the apatite  $c$ -axis length increased from  $\sim 135$  to 195  $\text{\AA}$ . Because component B is largest for the apatite mineral of smallest crystal size, this element of the contour presumably arises from a disordered phosphate phase not located in the crystalline environment.

The frequency of component A of the 900–1,200- $\text{cm}^{-1}$  contour increases from  $\sim 1,015$  to 1,030  $\text{cm}^{-1}$ , as crystal size of the poorly crystalline HA increases from  $\sim 125$  to 190  $\text{\AA}$ . It is probable that this component, together with component C centered near 1,105  $\text{cm}^{-1}$ , represent two elements of the  $\nu_3$  mode, whose degeneracy is lifted by the local symmetry of the HA crystal lattice. A previous study of apatite minerals, using Fourier self-deconvolution techniques, has attributed a 1,020- $\text{cm}^{-1}$  band to nonstoichiometric apatite containing  $\text{HPO}_4^{2-}$  and  $\text{CO}_3^{2-}$ , and an 1,125- $\text{cm}^{-1}$  band to stoichiometric apatite (Rey et al., 1991). These results are in agreement with the present study which indicates that the frequency of component A is reduced for less crystalline apatite minerals.

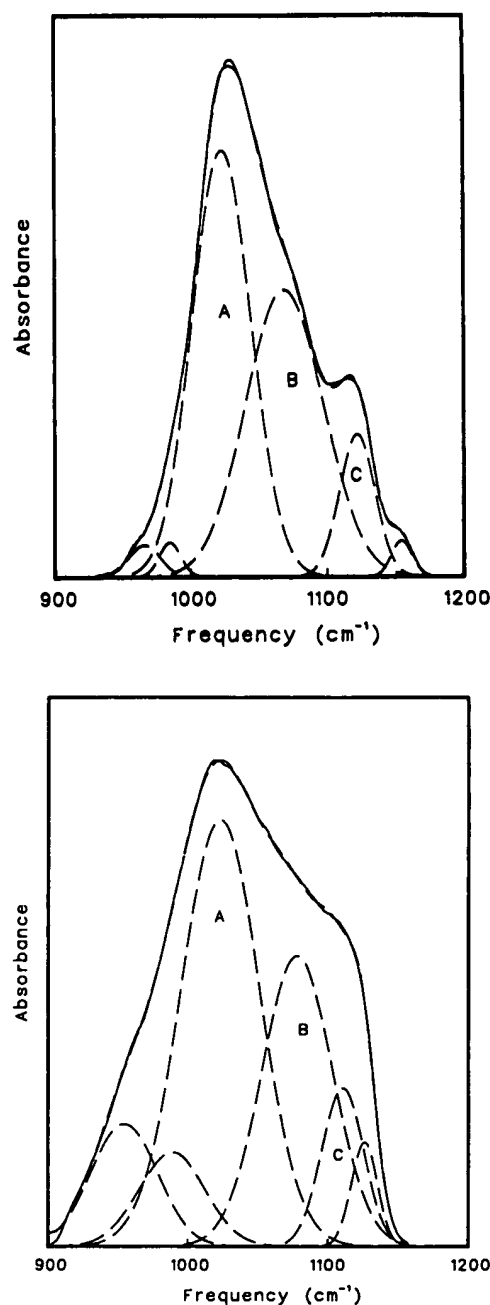


FIGURE 8 (A) FT-IR microscopy spectrum, 30  $\mu$  diameter region, taken from a region of calcified cartilage in the rat femur epiphyseal growth plate (—), with an overlay of the calculated contour and the individual components (---), as determined by a curve-fitting analysis. Using the percentage area component B, 38.6%, and Eq. 1, the crystal size of the apatite mineral was estimated to be 141  $\text{\AA}$ . (B) FT-IR microscopy spectrum, 30  $\mu$  diameter region, taken from a region  $\sim 40 \mu$  below that of part A (—), with an overlay of the calculated contour and the individual components (---), as determined by a curve-fitting analysis. Eq. 1 and percentage area component B, 26.3%, were used to estimate the crystal size of the mineral in this region (150  $\text{\AA}$ ).

A contribution from the phosphate ion of  $\text{HPO}_4^{2-}$  in the 900–1,200- $\text{cm}^{-1}$  absorbance of HA is also possible. Many studies have identified  $\text{HPO}_4^{2-}$  in biological and synthetic apatites, either incorporated into the crystal lattice, or on the surface of the HA (Arends and Davidson, 1975; Berry, 1967; Termine and Eanes, 1972). The free  $\text{HPO}_4^{2-}$  ion has  $C_{3v}$  symmetry, leading to eight normal modes of vibration (Table 1). The symmetric  $\nu_2$  A mode and the doubly degenerate  $\nu_6$  E P-O stretch appear in the 900–1,200- $\text{cm}^{-1}$  region, at 988 and 1,076  $\text{cm}^{-1}$ , respectively. When this ion is incorporated into a crystal lattice, as in brushite,  $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$ , the  $\nu_2$  mode appears as a doublet, at 987 and 1,000  $\text{cm}^{-1}$ , and the  $\nu_6$  mode appears between 1,058 and 1,137  $\text{cm}^{-1}$  (Berry and Baddiel, 1967). Accordingly, it is possible that two of the components of the 900–1,200- $\text{cm}^{-1}$  region obtained by the curve fit analysis arise from  $\text{HPO}_4^{2-}$  modes, indicating incorporation of the acid phosphate in the apatite lattice, or on its surface. The high frequency component near 1,140  $\text{cm}^{-1}$ , and the component near 989  $\text{cm}^{-1}$  may both arise from  $\text{HPO}_4^{2-}$ .

The carbonate ion may incorporate into the HA lattice at two different positions, known as the "A" (hydroxyl substitution) and "B" (phosphate substitution) sites (Bonel, 1972a, b; Elliot, 1961; Emerson and Fischer, 1962). Much work has been carried out to determine the effects of  $\text{CO}_3^{2-}$  substitution on the infrared spectra of apatites (Blumenthal et al., 1975; Elliot et al., 1985; LeGeros et al., 1968). It is known that the  $\nu_3$  absorbance band is broadened by incorporation of carbonate into the HA lattice. Additionally, the phosphate environment of type B carbonate-apatites is probably altered, leading to more than one type of phosphate absorbance from the apatite. However, it was not possible to correlate carbonate substitution with any of the components of the 900–1,200- $\text{cm}^{-1}$  region.

Fluoride is incorporated into the apatite lattice by means of substitution for the hydroxyl ion, causing an increase in crystal size by virtue of better packing of the crystal lattice (Posner et al., 1963; Schraer et al., 1962; Zipkin et al., 1962). Comparison of the fluoride substituted apatites to the unsubstituted apatites at equivalent

reaction time points shows the fluorapatites to have a slightly larger crystal size (Fig. 5A).

A relationship was established between the percentage area of the phosphate  $\nu_1$  mode, and the crystal size of the well crystallized apatite analogues (Fig. 7). The origin of the additional components needed to adequately fit for this series of compounds is unknown, but it is reasonable to assume that coupled corresponding motions of phosphate ions in the apatite lattice contribute to the absorbances in the 900–1,200- $\text{cm}^{-1}$  region. Although it is unclear why the  $\nu_1$  absorbance decreases with increasing crystallinity of the well crystallized apatites, the relationship is empirically useful for determining the crystal size of compounds in the 200–450-Å region.

This spectroscopic method for determining the crystal size of HA analogues is valuable for experimental conditions where x-ray diffraction experiments cannot be performed. It is noted that the two methods do not provide exactly the same type of molecular information. However, the FT-IR data from the biological specimens agrees with previously reported crystallite size determination for comparable samples (Boskey and Marks, 1985).

IR spectroscopy and x-ray diffraction are thus both suited to determine crystal sizes greater than 100 Å, and should be used in conjunction when possible to obtain the maximum amount of structural information. When x-ray diffraction of apatites cannot be performed, as is the case with thin, calcified, spatially inhomogeneous tissue sections, or in the case of poorly crystalline substances, the method developed here is valuable for assessing the crystallinity of HA phases.

## CONCLUSIONS

A novel infrared spectroscopic method has been developed to assess the crystallinity of poorly crystalline (crystal sizes of 100–190 Å) and well crystallized (200–450 Å) HA analogues, based on changes in the phosphate  $\nu_1$  and  $\nu_3$  absorbance. The technique is appropriate for the analysis of apatite minerals in both macroscopic samples and in microtomed calcified tissue sections. A preliminary application to the determination of the crystal size of apatite in a thin rat femur has been demonstrated.

The authors thank Dr. Steven Doty and Mr. Michael Maresca for their technical assistance.

Supported by National Institutes of Health grant DE-04141 and the Busch Memorial Fund of Rutgers University.

Received for publication 20 March 1991 and in final form 21 June 1991.

TABLE 1  $\text{HPO}_4^{2-}$  vibrational modes\*

P-O stretch	$\nu_2$ A	988
O-P-O bend	$\nu_3$ E	394
P-O(H) stretch	$\nu_3$ A	862
P-O stretch	$\nu_6$ E	1076
O-P-O bend	$\nu_4$ A	586
O-P-O bend	$\nu_5$ E	537
(P-)O-H stretch	$\nu_1$ A	2900
P-O-H bend	$\nu_3$ E	1230

\*Bailey and Holt, 1989.

## REFERENCES

- Arends, J., and C. L. Davidson. 1975.  $\text{HPO}_4^{2-}$  content in enamel and artificial carious lesions. *Calcif. Tiss. Res.* 18:65-79.
- Baddiel, C. B., and E. E. Berry. 1966. Spectra-structure correlations in hydroxy and fluorapatite. *Spectrochim. Acta.* 22:1407-1416.
- Bailey, R. T., and C. Holt. 1989. Fourier transform infrared spectroscopy and characterisation of biological calcium phosphates. In *Calcified Tissues*. D. W. L. Hukins, editor. MacMillan Press, Houndmills, Basingstoke, Great Britain. 93-120.
- Baud, C. A., J. M. Very, and B. Courvoisier. 1988. Biophysical study of bone mineral in biopsies of osteoporotic patients before and after long-term treatment with fluoride. *Bone.* 9:361-365.
- Berry, E. E. 1967. The structure and composition of some calcium-deficient apatites. *J. Inorg. Nucl. Chem.* 29:317-327.
- Berry, E. E., and C. B. Baddiel. 1967. The infra-red spectrum of dicalcium phosphate dihydrate (brushite). *Spectrochim. Acta.* 23a:2089-2097.
- Bhatnagar, V. M. 1968. Infrared spectra of hydroxyapatite and fluorapatite. *Bull. Soc. Chim. France.* 1771-1773.
- Blumenthal, N. C., F. Betts, and A. S. Posner. 1975. Effect of carbonate and biological macromolecules on formation and properties of hydroxyapatite. *Calcif. Tiss. Res.* 18:81-90.
- Bonel, G. 1972a. Contribution a l'etude de la carbonation des apatites. Part I. *Ann. Chim.* 7:65-87.
- Bonel, G. 1972b. Contribution a l'etude de la carbonation des apatites. Parts II and III. *Ann. Chim.* 7:127-144.
- Boskey, A. L., and S. C. Marks, Jr. 1985. Mineral and matrix alterations in the bones of incisors-absent (*ia/ia*) osteopetrotic rats. *Calcif. Tissue Int.* 37:287-292.
- Boskey, A. L., and A. S. Posner. 1973. Conversion of amorphous calcium phosphate to microcrystalline hydroxyapatite. A pH-dependent, solution-mediated, solid-solid conversion. *J. Phys. Chem.* 77:2313-2317.
- Cant, N. W., J. A. S. Bett, G. R. Wilson, and W. K. Hall. 1971. The vibrational spectrum of hydroxyl groups in hydroxyapatites. *Spectrochim. Acta.* 27A:425-439.
- Conway, E. J. 1957. *Microdiffusion Analysis and Volumetric Error*. Lockwood & Son Ltd., London.
- Cullity, B. D. 1956. *Elements of X-Ray Diffraction*. Addison-Wesley Publishing Company, Inc., Reading, MA. 97-99.
- Eanes, E. D., J. D. Termine, and M. U. Nysten. 1973. An electron microscopic study of the formation of amorphous calcium phosphate and its transformation to crystalline apatite. *Calcif. Tiss. Res.* 12:143-158.
- Elliot, J. C., D. W. Holcomb, and R. A. Young. 1985. Infrared determination of the degree of substitution of hydroxyl by carbonate ions in human dental enamel. *Calcif. Tissue Int.* 37:372-275.
- Elliott, J. C. 1961. The infrared spectrum of the carbonate ion in carbonate-containing apatites. (Abstr.) *J. Dent. Res.* 30:1284.
- Emerson, W. H., and E. E. Fischer. 1962. The infra-red absorption spectra of carbonate in calcified tissues. *Arch. Oral Biol.* 7:671-683.
- Fowler, B. O. 1974a. Infrared studies of apatites. I. Vibrational assignments for calcium, strontium, and barium hydroxyapatites utilizing isotopic substitution. *Inorg. Chem.* 13:194-207.
- Fowler, B. O. 1974b. Infrared studies of apatites. II. Preparation of normal and isotopically substituted calcium, strontium, and barium hydroxyapatites and spectra-structure-composition correlations. *Inorg. Chem.* 13:207-214.
- Fowler, B. O., E. C. Moreno, and W. E. Brown. 1966. Infra-red spectra of hydroxyapatite, octacalcium phosphate and pyrolysed octacalcium phosphate. *Arch. Oral Biol.* 11:477-492.
- Gron, P., H. G. McCann, and D. Bernstein. 1966. Effect of fluoride on human osteoporotic bone mineral. *J. Bone and Joint Surg.* 48A:892-897.
- Hasvold, Ö., and S. Dahm. 1977. The effect of fluoride on the transformation of amorphous calcium phosphate into crystalline apatite. *Calcif. Tiss. Res.* 22 (Suppl):425-427.
- Kragstrup, J., Z. Shijie, L. Mosekilde, and F. Melsen. 1989. Effects of sodium fluoride, vitamin D, and calcium on cortical bone remodeling in osteoporotic patients. *Calcif. Tissue Int.* 45:337-341.
- LeGeros, R. Z., O. R. Trautz, J. P. LeGeros, and E. Klein. 1968. Carbonate substitution in the apatite structure (1). *Bull. Soc. Chim. France.* 1712-1717.
- LeGeros, R. Z. 1981. Apatites in Biological Systems. *Prog. Crystal Growth Charact.* 4:1-45.
- Posner, A. S., F. Betts, and N. C. Blumenthal. 1979. Bone mineral composition and structure. In *Skeletal Research*. D. J. Simmons and A. S. Kunin, editors. Academic Press, New York. 167-192.
- Posner, A. S., E. D. Eanes, R. A. Harper, and I. Ziplit. 1963. X-ray diffraction analysis of the effect of fluoride on human bone apatite. *Arch. Oral Biol.* 8:549-570.
- Rey, C., M. Shimizu, B. Collins, and M. J. Glimcher. 1991. Resolution enhanced fourier transform infrared spectroscopy study of the environment of phosphate ion in the early deposits of a solid phase of calcium phosphate in bone and enamel and their evolution with age. 2. Investigations in the  $\nu_3$   $\text{PO}_4$  domain. *Calcif. Tissue Int.* In press.
- Rey, C., M. Shimizu, B. Collins, and M. J. Glimcher. 1990. Resolution-enhanced fourier transform infrared spectroscopy study of the environment of phosphate ions in the early deposits of a solid phase of calcium-phosphate in bone and enamel, and their evolution with age. 1. Investigations in the  $\nu_4$   $\text{PO}_4$  domain. *Calcif. Tissue Int.* 46:384-394.
- Schraer, H., A. S. Posner, R. Schraer, and I. Zipkin. 1962. The effect of fluoride on bone "crystallinity" in the growing rat. *Biochim. Biophys. Acta.* 64:565-567.
- Stutman, J. M., J. D. Termine, and A. S. Posner. 1965. Vibrational spectra and structure of the phosphate ion in some calcium phosphates. *Trans. NY Acad. Sci.* 27:669-675.
- Termine, J. D., and E. D. Eanes. 1972. Comparative chemistry of amorphous and apatitic calcium phosphate preparations. *Calcif. Tiss. Res.* 10:171-197.
- Termine, J. D., and D. R. Lundy. 1973. Hydroxide and carbonate in rat bone mineral and its synthetic analogues. *Calcif. Tiss. Res.* 13:73-82.
- Termine, J. D., and A. S. Posner. 1966. Infra-red determination of the percentage of crystallinity in apatitic calcium phosphates. *Nature (Lond.)* 211:268-270.
- Thompson, D. D., A. S. Posner, W. S. Laughlin, and N. C. Blumenthal. 1983. Comparison of bone apatite in osteoporotic and normal eskimos. *Calcif. Tissue Int.* 35:392-393.
- Warshawsky, H. 1982. Embryology and development of the skeletal system. In *The Musculoskeletal System Embryology, Biochemistry, and Physiology*. R. L. Cruess, editor. Churchill Livingstone, New York. 33-56.
- Ziplin, I., A. S. Posner, and E. D. Eanes. 1962. The effect of fluoride on the x-ray-diffraction pattern of the apatite of human bone. *Biochim. Biophys. Acta.* 59:255-258.